Enhanced HER2-dependent Immune Activation by Margetuximab, an Investigational Fc-engineered anti-HER2 mAb, Supports Combination with Checkpoint Blockade



Abstract

Background: Margetuximab (M) has demonstrated anti-tumor activity in patients with advanced HER2+ gastric cancer and PFS superiority to trastuzumab (T) in pre-treated metastatic HER2+ breast cancer patients¹. Similar to T, M inhibits HER2 signaling; additionally, M enhances engagement with the activating Fcy receptor (FcR)IIIA, while diminishing interaction with the inhibitory FcRIIB. Given the role of FcRs in coupling innate and adaptive immune responses, clinical studies of M in combination with PD-1 or PD-1 plus LAG-3 blockade have been initiated^{2,3}. The combination of M with tebotelimab, an investigational bispecific DART[®] molecule blocking PD-1 and LAG-3, has demonstrated encouraging early evidence of anti-tumor activity in patients with advanced HER2+ malignancies, including those with PD-L1-negative tumors³. Here we present data supporting the potential mechanisms underlying the anti-tumor activity elicited by M and tebotelimab combination therapy.

Methods: PBMCs were challenged with HER2-expressing tumor cell lines in the presence of M or T. Immune modulation was assessed by analysis of immune cell phenotype, gene expression profiles, and cytokine secretome. Functional activity of immune cells following exposure to the combination of M with tebotelimab was evaluated.

Results: M induced greater upregulation relative to T of both co-stimulatory (eg 4-1BB) and/or checkpoint molecules (eg PD-L1 and LAG-3) on NK cells and/or monocytes in PBMCs cocultured with HER2-expressing tumor cells. Compared with T, M also induced greater proinflammatory cytokine release, including IFNy that in turn upregulates PD-L1 and MHC class I expression on co-cultured tumor cells. These effects were not observed with an Fc-null version of M or were inhibited by the addition of FcR-blocking Abs, demonstrating Fc:FcR dependency of M-induced immune activation. Interestingly, CD8 T cells, which lack FcR, also exhibited LAG-3 upregulation in the above cultures³, suggesting the effects of M can also be mediated via a cascade mechanism. An IFNy neutralizing mAb reduced tumor cell-associated PD-L1 and MHC class I upregulation, but not LAG-3 induction on T-cells. PBMCs, pre-treated with a combination of M and tebotelimab in the presence of HER2-expressing tumor cells, demonstrated enhanced cytolytic activity against both the canonical NK cell target, K562, or M-opsonized HER2-positive tumor cells.

Conclusions: M, through an Fc-dependent mechanism, can upregulate checkpoint molecules on NK-cells, CD8 T cells and tumor cells in vitro, thereby sensitizing them to immune checkpoint blockade. In turn, PD-1 and LAG-3 blockade by tebotelimab enhances M-mediated NK cell cytolytic activity in vitro.

Introduction

Margetuximab

anti-HER2

Tebotelimab

anti-PD-1 x anti-LAG-3

Engineered IgG1 Fc

lgG4 Fc

- Margetuximab: Fc-engineered anti-HER2 therapeutic mAb - Engineering
 - Fab: Same specificity and comparable affinity as trastuzumab - Fc: Increased affinity for both allelic variants of the activating FcyRIIIA (CD16A) compared to Trastuzumab; Binding to the low-affinity CD16A-158F variant by margetuximab exceeds that of trastuzumab for the high-affinity CD16A-158V variant; decreased affinity for the inhibitory FcyRIIB (CD32B)
- Bioactivity
- Enhanced in vitro ADCC compared to trastuzumab
- Clinical studies
- SOPHIA: Phase 3 study of margetuximab plus chemotherapy compared to trastuzumab plus chemotherapy in patients with metastatic HER2-positive breast cancer who had received prior chemotherapy and at least one form of anti-HER2 treatment. Improved PFS compared to control arm¹
- MAHOGANY: Phase 2/3 study in patients with HER2-positive gastric cancer in first-line therapy in combination with PD-1 blockade²
- **Tebotelimab:** PD1 x LAG-3 bispecific IgG4 DART protein

- Engineering

- Tetravalent (bivalent for each target) IgG4 DART molecule - Bioactivity
- Binds to and blocks PD-1 and LAG-3 concomitantly or independently - Greater synergistic in vitro T-cell activation (IFNy release) compared to the combination
- of its individual constituents
- Clinical study
- Anti-tumor activity in various advanced solid tumors³
- Margetuximab and Tebotelimab combination therapy
- Evidence of anti-tumor activity in patients with advanced HER2+ malignancies, including those with PD-L1-negative tumors⁴



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Transcription profiles (RNA-seq) of margetuximab (M) or trastuzumab (T)-treated PBMCs from two healthy donors in the presence of HER2-expressing N87 cells (E:T ratio = 15:1, 48hr incubation; Donor 1: CD16A 158 V/F; Donor 2: CD16A 158 F/F). **A.** Immune-related genes with \geq 2-fold increase following M or T treatment were grouped as indicated (Log₂ normalized) expression values [DEseq]). **B.** Fold increase of *IFNG, GZMB, CD274* and *LAG3* gene expression.

Margetuximab-mediated Engagement With **HER2-positive Tumor Cells in Vitro Induces Co-signaling Molecules on Effector Cells**

Induction of both checkpoint (PD-L1 and LAG-3) and co-stimulatory (CD137) pathways Margetuximab mediates greater activity relative to trastuzumab



*p<0.05, ** p<0.01 (two-tailed paired Student's t-tes

Cell surface expression of PD-L1, LAG-3 and CD137 (4-1BB) were assessed by FACS analysis of PBMCs treated by margetuximab (M) or trastuzumab (T) in the presence of HER2 expressing N87 gastric cancer cells (A) or SKBR3 breast cancer cells (B). Data are shown as mean +/- SEM of 7 separate experiments.

Results

Margetuximab-induced PD-L1 or LAG-3 Upregulation on NK Cells and Monocyte Depends upon the **Engagement of CD16A**



NK cells and monocytes from PBMCs treated with margetuximab upregulate PDL-1 and LAG-3, which was prevented by CD16A-blockade. An aglycosylated Fc-null version of margetuximab failed to upregulate PD-L1 or LAG-3. Notably, CD32 blockade prevented LAG-3 upregulation by monocytes.

Margetuximab Induces Enhanced PD-L1 and MHC Class I Expression on Tumor Cells via an **IFNy-dependent** Mechanism



Healthy donor PBMCs were treated with margetuximab (M) or trastuzumab (T) in the presence of HER2-expressing tumor cells for 3 days. (A) Levels of cytokines in culture supernatant was assessed using Milliplex[®] MAP human TH17 cytokine panel. Data are shown as mean +/- SD of 4 separate experiments. Out of 11 cytokines, IFNy, TNFα, IL-6, and IL-1β showed significant increase post M or T treatment. Differences in cytokine release were not significant, except for TNF α (*p < 0.05, two-tailed paired Student's t-test). (B) Supernatant (sup) was collected to treat N87 or SKBR3 cells for 24 hours in the presence or absence of anti-IFNy-blocking Ab. The expression of PD-L1 and HLA-ABC on the surface of tumor cells was assessed by flow cytometry. Data are from a representative experiment of 3 performed.



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Healthy donor PBMCs were pre-treated with margetuximab ± tebotelimab in the presence of HER2-expressing N87 gastric cancer cells for 6 days, as indicated. **A.** Natural killer cell activity (NK Activity) was assessed by FACS analysis of annexin V/ 7-AAD staining of K562 target cells. Percent of cytotoxicity represents sum of % apoptotic cells Annexin V+7-AAD-) and % dead cells (Annexin V+7-AAD+). **B.** Antibody-dependent cell cytotoxicity (ADCC) was assessed by measuring the luminescence of luciferase-expressing, margetuximab-opsonized SKBR3 cells. Cytotoxicity was normalized by setting no treatment control as 0.

Conclusions

Engagement of HER2+ tumor cells with margetuximab induces:

- Enhanced inflammatory genes expression, upregulation of the co-inhibitory molecules, PD-L1 and LAG-3, and expression of the co-stimulatory molecule, CD137 (4-1BB) by immune effector cells relative to trastuzumab
- Induction of LAG-3 and PD-L1 on NK cells and monocytes is mediated by Fc:CD16A interaction
- Enhanced PD-L1 and MHC class I expression on tumor cells, mediated by IFNy
- Exposure of margetuximab-targeted PBMC to tebotelimab, a bispecific DART molecule that blocks PD-1 and LAG-3, may prime NK cells to enhance their intrinsic cytolytic capacity

Acknowledgements

1. Rugo et al., 2021 JAMA Oncology doi:10.1001/jamaoncol.2020.7932. **2.** Catenacci et al., 2020 Lancet Oncology 21(8):1066-76. **3.** Luke et al., 2020 ASCO oral presentation. **4.** Patel et al., 2020 SITC poster presentation.